

RESEARCH

Open Access

A paradigm of fragile Earth in Priestley's bell jar

Daniel Martin^{1,2*}, Andrew Thompson³, Iain Stewart⁴, Edward Gilbert¹, Katrina Hope⁵, Grace Kawai¹ and Alistair Griffiths⁶

Abstract

Background: Photosynthesis maintains aerobic life on Earth, and Joseph Priestley first demonstrated this in his eighteenth-century bell jar experiments using mice and mint plants. In order to demonstrate the fragility of life on Earth, Priestley's experiment was recreated using a human subject placed within a modern-day bell jar.

Methods: A single male subject was placed within a sealed, oxygen-depleted enclosure (12.4% oxygen), which contained 274 C₃ and C₄ plants for a total of 48 h. A combination of natural and artificial light was used to ensure continuous photosynthesis during the experiment. Atmospheric gas composition within the enclosure was recorded throughout the study, and physiological responses in the subject were monitored.

Results: After 48 h, the oxygen concentration within the container had risen to 18.1%, and hypoxaemia in the subject was alleviated (arterial oxygen saturation rose from 86% at commencement of the experiment to 99% at its end). The concentration of carbon dioxide rose to a maximum of 0.66% during the experiment.

Conclusions: This simple but unique experiment highlights the importance of plant life within the Earth's ecosystem by demonstrating our dependence upon it to restore and sustain an oxygen concentration that supports aerobic metabolism. Without the presence of plants within the sealed enclosure, the concentration of oxygen would have fallen, and carbon dioxide concentration would have risen to a point at which human life could no longer be supported.

Keywords: Atmosphere, Carbon dioxide, Oxygen, Hypoxia, Photosynthesis, Plants

Background

The Earth supports a fragile ecosystem, and its inhabitants depend for their survival upon complex interactions between them, which have developed over billions of years. Imbalance of one component in this bionetwork can have far-reaching effects on organisms whose existence relies upon the presence of other species. Despite the ability to alter their environment in diverse ways, humans are reliant for their survival upon an element derived primarily from plants and produced by chlorophyll during photosynthesis, oxygen (O₂).

Photosynthesis is arguably the single most important chemical process on our planet, and the first colour images captured of Earth from space revealed the vast green hues

of the landmasses supporting plant life, confirming its dominance within our ecosystem. Using energy from sunlight, chlorophyll strips electrons from water molecules, which then convert atmospheric carbon dioxide (CO₂) into carbon compounds, producing O₂ as a byproduct. Whilst mechanisms that use alternative naturally available compounds to release energy exist, the abundance of water on the surface of the Earth meant that photosynthesis rapidly became the foremost bio-energetic pathway on the planet. During the early era of chlorophyll photosynthesis, approximately 2,400 million years ago [1], the atmosphere was rich in CO₂, whilst O₂ was scarce. As time progressed and photosynthetic species slowly overwhelmed the surface of the Earth, the concentration of O₂ rose and eventually reached levels we are accustomed to today.

In the early 1770 s, Joseph Priestley conducted a series of experiments that led to the discovery of the intimate relationship between plant and animal life [2]. In his principal experiment, Priestley placed a mouse within a sealed jar and observed it to eventually perish. When repeated with

* Correspondence: daniel.martin@ucl.ac.uk

¹UCL Centre for Altitude, Space and Extreme Environment Medicine, Portex Unit, Institute of Child Health, 30 Guilford Street, London WC1N 1EH, UK

²Division of Surgery and Interventional Science, University College London, 9th Floor, Royal Free Hospital, London NW3 2QG, UK

Full list of author information is available at the end of the article

sprigs of mint within the jar, neither did the animal die 'nor was it at all inconvenient to a mouse' [2]. He had made the breakthrough that plants produce a substance which is life-giving to animals and then went on to describe 'dephlogisticated air', which, thanks to the French chemist Antoine Lavoisier, soon became known as 'oxygen'. The story of photosynthesis was completed in 1779 when a Dutchman, Jan Ingenhousz, demonstrated that the process by which plants produce O₂ is dependent upon light.

We hypothesised that a human could survive within a sealed modern-day bell jar, even if the O₂ concentration within was significantly reduced from the outset, provided that it contained sufficient plant matter to generate O₂ and remove CO₂ via photosynthesis.

Methods

Formal ethical approval was not sought for this experiment as it was designed for the purpose of a television demonstration; consent was implied through the subject's involvement in the project and participation in the event. The Chair of the University College London Committee on the Ethics of Non-NHS Human Research approved this strategy. Prior to commencing the experiment, a full medical screening questionnaire was completed by the subject, and he was assessed by a physician with experience in high altitude and acute hypoxia research (DM). The protocol was explained to the subject in full, along with a description of the potential risks and safety measures in place. A standard resuscitation kit was available throughout the experiment, along with bottled supplemental oxygen. A physician trained in Advanced Life Support was also present outside the container throughout the experiment, with the ability to enter the container at any point should there be concerns regarding the welfare of the subject.

We constructed the first human recreation of Priestley's 'mouse in a bell jar' experiment to demonstrate the ability of plants to generate sufficient O₂ to sustain human life in an enclosed environment [3]. A healthy 47-year-old male was placed within a transparent airtight container measuring 2.0 × 2.5 × 6.0 m (30 m³, Figure 1), itself placed within the rainforest biome at the Eden Project, Cornwall, UK. A selection of plants known for their high photosynthetic yield (under certain environmental conditions) was placed within the container. Prior to the experiment, containerised plants were grown in a peat-free Eden Project Melcourt mix within a standard glasshouse at a relative humidity of 70% to 80% and temperature range of 15°C to 30°C. During this growing phase, the plants were watered with liquid nutrient feed at 20 ml/L (N 177 ppm, P 35 ppm, K 119 ppm, Ca 49 ppm, Mg 17 ppm, B 0.2 ppm, Cu 0.08 ppm, Fe 1.44 ppm, Mn 0.48 ppm, Mo 0.04 ppm and Zn 0.64 ppm). In total, 274 plants consisting of 18 different taxa were placed within the container, with 10,967 leaves (excluding *Tillandsia usneoides*) and a total leaf area of 1,106,033 cm²

(Table 1). A mixture of C₃ (ribulose diphosphate carboxylase utilising) and C₄ (phosphoenolpyruvate carboxylase utilising) plants were selected in order to maximise photosynthetic potential within the container. Several C₄ carbon fixation plants were grown, including *Miscanthus x giganteus* and *Zea mays* (maize), because they have advantages over C₃ plants, resulting in superior carbon-gaining capacities and photosynthetic efficiency [4]. During the experiment, the subject regularly irrigated the plants when deemed necessary from a water source within the container.

In order to more clearly demonstrate oxygen production and highlight the effectiveness of photosynthesis in preserving human life, the environment within the container was rendered hypoxic at the start of the experiment. Three hypoxic generators (Hypoxico Everest Summit II, Hypoxico Inc, New York, NY, USA) were used to reduce the concentration of O₂ in the container. These devices consist of a molecular sieve system that uses zeolite to separate nitrogen from O₂ in the air and consequently provides a nitrogen-rich gas mixture to purge the atmosphere within the container. Connected to the container, and in conjunction with a one-way pressure relief valve, the hypoxic generators reduced the concentration of O₂ to 12.4% prior to commencing the experiment. Once the subject was sealed inside the container and safety procedures had been confirmed, the hypoxic generators were switched off and the one-way valves were closed. Artificial lighting (8 × 2,000 W systems; ARRI, Munich, Germany) was placed around the container externally and switched on at the beginning of the experiment. A split air-conditioning unit (Clima 16 HP Portable Air Conditioner, Toshiba, Tokyo, Japan) was used to maintain temperatures for optimal plant growth and comfort for the subject whilst ensuring a sealed atmosphere. The concentrations of O₂ and CO₂ within the container were monitored with a gas analyser (Aspida, Analox, London, UK) and plotted every hour along with temperature and humidity from a digital hygrothermometer (Brannan, Cumbria, UK). The subject's heart rate and arterial O₂ saturation (SpO₂) were monitored continuously (Johnson and Johnson Dinamap MPS Monitor and Onyx 9500, Nonin, Plymouth, MN, USA); respiratory rate was recorded hourly by manual calculation.

Results

The concentration of O₂ in the container rose throughout the experiment, peaking at 18.1% in the final hour (hour 48; Figure 2). The CO₂ concentration fluctuated depending on the subject's activity within the container (declining noticeably during sleep), but there was an overall rise that peaked at 0.66%, approximately halfway through the experiment (Figure 3). There was a diurnal variation in temperature (25.3°C to 28.4°C), and humidity varied between 57% and 87%. On entering the hypoxic



Figure 1 The sealed container with plants, the subject and external artificial lighting.

container, the subject had a heart rate of 90 beats per minute, respiratory rate of 20 breaths per minute and SpO₂ of 86%. These figures returned to the subject's resting normal values as the concentration of O₂ rose within the container. The subject's final SpO₂ was 99% (Figure 4).

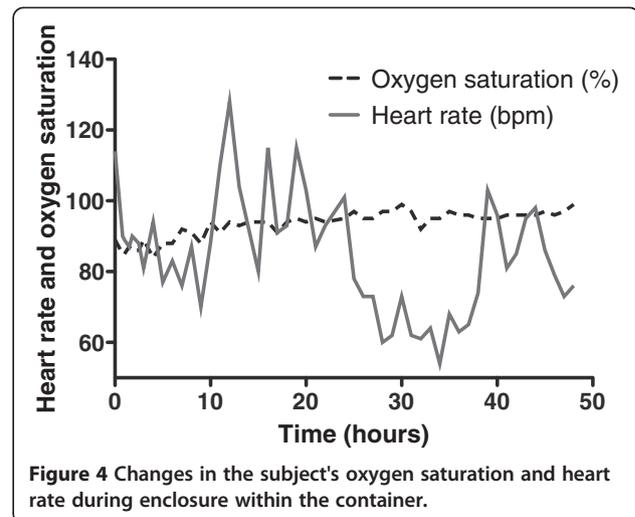
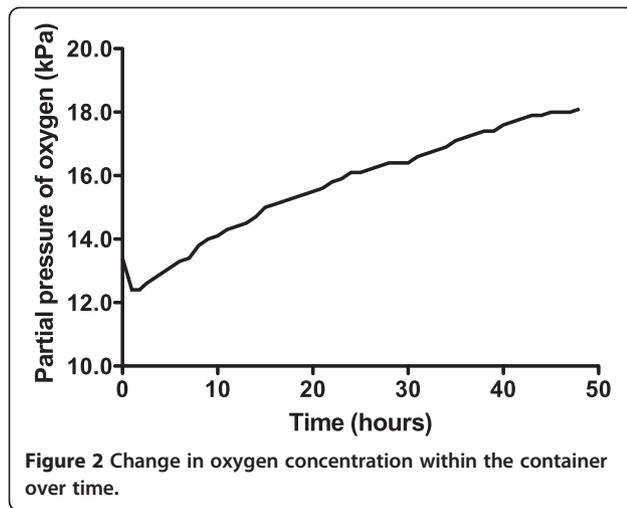
Discussion

The design of the biological ecosystem in this study was such that human life was sustained for 48 h and the initial hypoxic environment restored to one of near-normal O₂ concentration. In the early 1990's, the 'Biosphere 2'

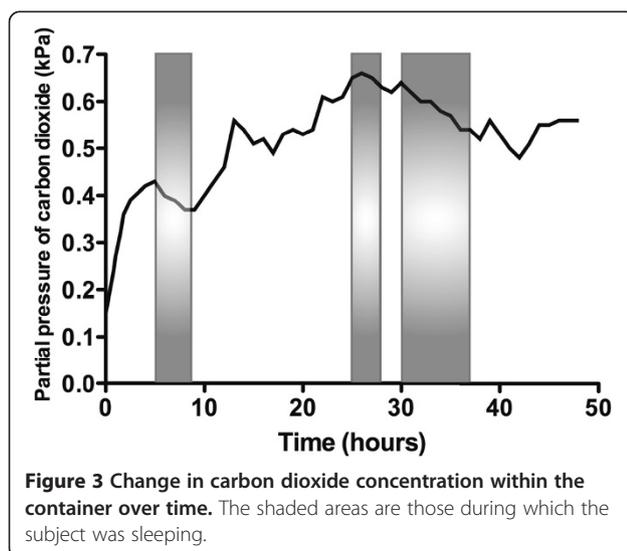
Table 1 Taxa, number of leaves and leaf area of the plants placed within the container

Scientific plant name	Common name	Number of plants	Total number of leaves	Total upper leaf area (cm ²) ^a
<i>Z. mays</i>	Corn or maize	25	375	37,192
<i>M. x giganteus</i>	Miscanthus	30	720	72,841
<i>Leymus arenarius</i>	Lyme grass	21	3,528	7,281
<i>Calathea rufibarba</i> 'Wavestar'	Calathea	12	1,560	126,204
<i>Ctenanthe</i> 'Golden Mosaic'	Ctenanthe	1	160	35,200
<i>Spathiphyllum</i> Cv.	Peace lily	3	282	214,656
<i>Saccharum officinarum</i>	Sugarcane	2	72	34,534
<i>Musa acuminata</i>	Banana	9	224	204,056
<i>Astelia chathamica</i> 'Silver Spear'	Astelia	33	1,188	26,479
<i>Furcraea longaevea</i>	Furcraea	40	400	12,301
<i>Anthurium andreanum</i> Cv.	Flamingo flower	2	24	163,296
<i>Chrysopogon zizanioides</i>	Vetiver	2	1,088	116,873
<i>Aechmea</i> Cv.	Bromeliad	14	280	20,475
<i>Eruca sativa</i>	Rocket	16	272	11,288
<i>Lactuca sativa</i>	Lettuce	16	272	11,288
<i>Mentha spicata</i>	Mint	5	450	1,512
Ferns	Ferns	9	72	10,557
Totals		274	10,967	1,106,033

Individual leaf areas were determined by tracing a leaf onto graph paper; the area of the petiole was not included within the calculations. From each individual plant, a subsample representing three small, three medium and three large leaves were harvested, and the mean of each of the three leaves was taken and used to provide a representative small, medium and large leaf area. The number of small, medium and large leaves in each individual plant was then counted, and the corresponding areas were used to estimate the total leaf surface area. ^aTotal leaf area was calculated as both the upper and lower sides of the leaves. *T. usneoides*, a moss, was also placed within the container, but it was not possible to calculate leaf area.



experiment was conducted to explore the feasibility of self-sustaining biospheres in space. This grand design consisted of a 200 m³ atmosphere within a dome that contained eight volunteers, which was designed to sustain them for 2 years [5]. However, the O₂ concentration within the biosphere dropped from 20.9% to 14.2% after 16 months, so additional O₂ had to be added to the atmosphere [6]. This decline was traced to a two-step process: firstly, there was O₂ loss to organic soil matter producing CO₂, and secondly, the CO₂ was being captured by structural concrete to form calcium carbonate [5]. In the current experiment, the initial O₂ concentration of 12.4% (equivalent to approximately 4,500 m above sea level) resulted in a marked reduction in the subject's SpO₂ and represents an acute hypoxic exposure that is frequently associated with symptoms of altitude-related illness [7]. During the last few hours of the study, there was a small reduction in rate of the O₂ concentration



rise, perhaps due to deterioration in the condition of the plants, noticeable towards the end of the experiment. Direct heating and excessive light exposure, arguably both present in this experiment, can lead to the denaturing of enzymes within chlorophyll [8]. There were fluctuations in CO₂ concentration throughout the study, with a tendency for it to rise as time progressed (Figure 3).

As well as providing an insight into the use of plants to maintain a self-sufficient biosphere, such as would be required on the surface of extra-terrestrial bodies without an atmosphere, our experiment highlights the detrimental effects of a markedly increased CO₂ concentration. CO₂ concentrations have altered dramatically over the course of the Earth's history [9], and there is much concern that levels are now rising at an alarming rate [10]. Under certain environmental conditions, increasing the ambient concentration of CO₂ can be beneficial, increasing photosynthetic activity, plant growth and yield [11,12]. Using CO₂ enrichment to increase plant growth and yield is now commonplace in commercial glasshouse crop production, with optimal levels being between 700 and 1,000 ppm [13]. However, in some species, super-elevated CO₂ concentrations (over 2,000 ppm) induces foliar symptoms of chlorosis and necrosis [14,15], and levels above 10,000 ppm are known to cause damage to young maize plants after 48 h in the form of 'yellow streaks' [16]. During this experiment, the CO₂ levels remained above 2,000 ppm and reached a maximum of 6,600 ppm, yet yellow streaks were observed on the maize plants by the end. It is possible that damage to the maize may have also reduced the photosynthetic yield and the production of O₂ towards the end of this experiment. This study, therefore, provides an insight into the use of plants to maintain a self-sufficient biosphere, such as would be required on the surface of extra-terrestrial bodies without an atmosphere, and the potentially detrimental effects of a dramatically increased CO₂ concentration.

Conclusions

This simple experiment is a humble reminder of the integral relationship between animal and plant life on Earth, in which the former owe their existence to the latter. Without the presence of plants within the sealed environment, the concentration of O₂ would have fallen and CO₂ concentration would have risen to a point at which human life could no longer be supported. Whilst O₂ sustains human life and plants maintain its level within the atmosphere with remarkable efficiency, the fundamental role of photosynthesis is arguably taken for granted. Deprived of plants, the subject within the container would have succumbed to the effects of severe hypoxaemia. The experiment reminds us of our total dependency upon plants, and the ecosystem in which they exist.

Competing interests

The author declares that they have no competing interests.

Authors' contributions

AT conceived the idea, and the experiment was designed by AT, AG and DM. The experiment was conducted by DM, KH, EG, GK, AT and IS. Data were analysed by DM, and the manuscript was written by DM, AG and EG. All authors discussed the results and implications and commented on the manuscript at all stages. All authors read and approved the final manuscript.

Acknowledgements

We would like to thank the BBC Scotland team for the 'How to Make a Planet' series and all the staff at the Eden Project in Cornwall who made this experiment possible.

Author details

¹UCL Centre for Altitude, Space and Extreme Environment Medicine, Portex Unit, Institute of Child Health, 30 Guilford Street, London WC1N 1EH, UK. ²Division of Surgery and Interventional Science, University College London, 9th Floor, Royal Free Hospital, London NW3 2QG, UK. ³BBC Television, Zone 2.20, BBC Pacific Quay, Glasgow G51 1DA, UK. ⁴School of Geography, Earth & Environmental Sciences, Plymouth University, Plymouth PL4 8AA, UK. ⁵Centre of Human & Aerospace Physiological Sciences, School of Biomedical Sciences, King's College London, London SE1 1UL, UK. ⁶The Eden Project, Bodelva, Cornwall PL24 2SG, UK.

Received: 14 March 2012 Accepted: 4 September 2012

Published: 4 September 2012

References

1. Blankenship RE: **Origin and early evolution of photosynthesis.** *Photosynth Res* 1992, **33**:91–111.
2. Priestley J: *Experiments and Observations on Different Kinds of Air.* London: W. Bowyer and J. Nichols; 1774.
3. : **Water the plant, take a breath.** *Science* 2011, **333**:1685.
4. Sage RF, Zhu XG: **Exploiting the engine of C(4) photosynthesis.** *J Exp Bot* 2011, **62**:2989–3000.
5. Allen J, Nelson M: **Overview and design biospherics and biosphere 2, mission one (1991–1993).** *Ecol Engin* 1999, **13**:15–29.
6. Dempster WF: **Biosphere 2. System dynamics and observations during the initial two-year closure trial.** In *23rd International conference on environmental systems: July 1993; Colorado.* Warrendale: SAE International; 1993.
7. Hackett PH, Roach RC: **High-altitude illness.** *N Engl J Med* 2001, **345**:107–114.
8. Bohning RH: **Time course of photosynthesis in apple leaves exposed to continuous illumination.** *Plant Physiol* 1949, **24**:222–240.
9. Pearson PN, Palmer MR: **Atmospheric carbon dioxide concentrations over the past 60 million years.** *Nature* 2000, **406**:695–699.
10. Alley R, Berntsen T, Bindoff NL, Chen Z, Chidthaisong AF P, Gregory J, Hegel G, Heimann M, Hewitson B: *Climate Change 2007: The Physical Science Basis,*

Summary for Policymakers. Geneva: Intergovernmental Panel on Climate Change Secretariat; 2007.

11. Körner C: **Biosphere responses to CO₂ enrichment.** *Ecol Appl* 2000, **10**:1590–1619.
12. Drake BG, Gonzalez-Meler MA, Long SP: **More efficient plants: a consequence of rising atmospheric CO₂?** *Annu Rev Plant Physiol Plant Mol Biol* 1997, **48**:609–639.
13. Hanan JJ: *Greenhouses. Advanced technology for protected horticulture.* London: CRC Press; 1998.
14. Ehret DL, Jolliffe PA: **Leaf injury to bean plant grown in carbon dioxide enriched atmospheres.** *Can J Bot* 1985, **63**:2015–2020.
15. van Berkel N: **Injurious effects of high CO₂ concentration on cucumber, tomato, chrysanthemum and gerbera.** *Acta Horticulturae* 1984, **162**:101–112.
16. Schwarz M: **Carbon toxicity in plants.** *Acta Horticulturae* 1999, **481**:685–688.

doi:10.1186/2046-7648-1-4

Cite this article as: Martin *et al.*: A paradigm of fragile Earth in Priestley's bell jar. *Extreme Physiology & Medicine* 2012 **1**:4.

Submit your next manuscript to BioMed Central and take full advantage of:

- **Convenient online submission**
- **Thorough peer review**
- **No space constraints or color figure charges**
- **Immediate publication on acceptance**
- **Inclusion in PubMed, CAS, Scopus and Google Scholar**
- **Research which is freely available for redistribution**

Submit your manuscript at
www.biomedcentral.com/submit

